NAPHTHOFURAN PRECURSORS OF SESQUITERPENOID ALDEHYDES IN DISEASED GOSSYPIUM*†

ROBERT D. STIPANOVIC, ALOIS A. BELL and CHARLES R. HOWELL

National Cotton Pathology Research Laboratory, P.O. Drawer JF, College Station, Texas 77840, U.S.A.

(Received 28 January 1975)

Key Word Index—Gossympium barbadense; Malvaceae; cotton; phytoalexins; 3,4-dihydroxy-5-isopropyl-7-methyl-2*H*-naphtho[1,8-bc]furan; 3-hydroxy-5-isopropyl-4-methoxy-7-methyl-2*H*-naphtho[1,8-bc]furan; vergosin; hemigossypol; 6-methoxyhemigossypol; *Verticillium dahliae*.

Abstract—Crude terpenoid fractions from stele tissues of *Gossypium barbadense* infected with *Verticillium dahliae* contained two prominent compounds that were nonaldehydic, but readily autooxidized to the terpenoid aldehydes hemigossypol and 6-methoxyhemigossypol. Spectral analyses, chemical studies, and comparisons with model compounds showed that these compounds were the sesquiterpenoids 3,4-dihydroxy-5-isopropyl-7-methyl-2*H*-naphtho[1,8-*bc*]furan (1a) and 3-hydroxy-5-isopropyl-4-methoxy-7-methyl-2*H*-naphtho[1,8-*bc*]furan (1b). The trivial names desoxyhemigossypol and desoxy-6-methoxyhemigossypol are proposed for (1a) and (1b). The sesquiterpenoid, previously isolated and named vergosin, is apparently desoxy-6-methoxyhemigossypol.

INTRODUCTION

The occurrence of antifungal substances in cotton plants (Gossypium hirsutum L. and G. barbadense L.) infected with Verticillium dahliae Kleb. is well documented [1-5]. These substances have generally been referred to as the gossypol-related phytoalexins. Zaki et al. [4,5] extensively analyzed two of these compounds. One was identified as (1,6,7-trihydroxy-5-isopropyl-3hemigossypol methyl-8-naphthaldehyde), and the second was purported to be 1-hydroxy-5-isopropyl-7-methoxy-3-methyl-8-naphthaldehyde (vergosin). We confirmed the structure of hemigossypol and isolated the related aldehydes 6-methoxy hemigossy-(1,7-dihydroxy-5-isopropyl-6-methoxy-3pol methyl-8-naphthaldehyde) and 6-deoxy hemigos-(1,7-dihydroxy-5-isopropyl-3-methyl-8naphthaldehyde) but failed to find an aldehyde corresponding with vergosin [3].

In our previous studies [3] we isolated two sesquiterpenoids that did not contain aldehyde

groups. Our present data indicate that these compounds are the 2*H*-naphtho[1,8-*bc*] furans (1a and 1b). Compound (1b) is identical to the compound previously reported, but incorrectly identified as vergosin [4,5].

RESULTS

Identification of desoxy-6-methoxyhemigossypol

Desoxy-6-methoxyhemigossypol (1b) was identified as 3-hydroxy-5-isopropyl-4-methoxy-7-methyl-2H-naphtho[1,8-bc]furan. The high-resolution MS of (1b) showed the molecular formula $C_{16}H_{18}O_3$ (found: m/e 258·126003, 100%; required: 258·125580). Major fragment ions were also noted at m/e 243 (80%, M^+ -Me) and 228 (26%, M^+ -2Me).

(1a) R = OH (1b) R = OMe

^{*}A preliminary report of this work was presented at the 29th Southwest Regional Meeting of the American Chemical Society, San Antonio, Texas, 5-7 December, 1973.

[†] For the previous paper in this series see Ref. [3].

Three fragment ions observed in the MS by Zaki et al. [4] were important in their analyses. In our analysis of (1b), the elemental composition of these three ions was determined by high resolution mass measurements. The ion at m/e 229 $(M^+-29, 8\%)$ had the formula $C_{14}H_{13}O_3$ (found: 229.085792: required: 229.086455) formed by the loss of C₂H₅, not CHO as previously suggested [4]. The ions at m/e 213 (4%) and 212 (4%) had the formulae $C_{13}H_9O_3$ (found: 213.055830): required: 213-055155) and C₁₄H₁₂O₂ (found: 212:083142; required: 212:083720), respectively. Thus, m/e 213 was formed by the loss of three Me groups, and m/e 212 resulted from the loss of a Me group and a OMe group. These ions are unrelated to any type of tautomerization between a peri hydroxyl and aldehyde as previously suggested [4]. Our IR of (1b) showed hydroxyl absorption at 3425 cm⁻¹, but there was no carbonyl peak at 1680 cm⁻¹.

The PMR of (1b) indicated the presence of an isopropyl group in which the chemical shift of both the methine proton (δ 3·70, 1H, sept) and the methyl protons (δ 1·50, 6H, d) were shifted down field, due to deshielding by the aromatic ring current. An aromatic Me group and a OMe group were indicated by three proton singlets at δ 2·40 and 3·73; two aromatic protons appeared as singlets at δ 6·35 and 7·10. A phenolic proton appeared as a broad peak center at δ 5·89, which disappeared on D₂O exchange and a two-proton singlet was observed at δ 5·61.

Assignment of the two-proton singlet at δ 5.61 to the aromatic methylene group was based on comparison with two model compounds. This singlet appears at δ 5.70 in 2H-naphtho[1,8-bc]furan [6], and at δ 5.79 in desoxygossypol tetramethyl ether [7]. 2H-Naphtho[1,8-bc]furan and the homocyclic analogue phenalene readily form stable delocalized radicals [6,8], which account for the rapid oxidation of (1b) in air, especially in the presence of UV light. The position of methoxylation in (1b) was confirmed by its photooxidation to give predominantly 6-methoxyhemigossypol.

Identification of desoxyhemigossypol

Desoxyhemigossypol (1a) was identified as 3,4-dihydroxy-5-isopropyl-7-methyl-2H-naphtho[1,8-bc]furan. The molecular formula $C_{15}H_{16}O_3$ was

confirmed by the high resolution MS (found: m/e 244·109651, 74%; required: 244·109930). The PMR spectrum showed signals for aromatic isopropyl (δ 1·47, 6H, d and δ 3·59. 1H, sept) and Me (δ 2·40, 3H, s) groups. Resonance signals for two phenolic protons appeared as a broad peak centered at δ 5·55. Two aromatic protons absorbed at δ 6·29 (s) and 6·97 (s). The methylene protons appeared as a sharp two-proton singlet at δ 5·57, superimposed on the broad phenol proton resonance.

The MS of (1a), like that of (1b), showed loss of Me (m/e 229, 100%), C_2H_5 (m/e 215, 12%), 2Me (m/e 214, 16%), and 3Me (m/e 199, 9%). The structure of (1a) was confirmed by its photooxidation to hemigossypol.

DISCUSSION

Desoxy-6-methoxyhemigossypol (1b) isolated in our studies appears to be identical to the compound previously isolated and identified as vergosin by Zaki et al. [4]. Both their data and ours show the same empirical formulae and similar UV, PMR, MS and chromatographic behavior. Compound (1b), like their compound, slowly turned pink when treated in light with acidic phloroglucinol, a reagent for aldehydes. This reaction was apparently preceded by autooxidation of (1b) to the aldehyde 6-methoxyhemigossypol.

Zaki et al. [4] concluded that their compound contained an aldehyde group because of three observations: (a) an IR absorption at $1680 \,\mathrm{cm}^{-1}$; (b) fragments in the MS at m/e 229, 213 and 212, that they attributed to loss of CHO, CHO₂ and CH₂O₂, respectively, from the M⁻; and (c) an NMR peak at δ 7·3 that they concluded was due to an aldehyde proton.

We found no evidence for an aldehyde group in (1b). No carbonyl absorption occurred at $1680 \, \mathrm{cm^{-1}}$ in freshly purified (1b), although some carbonyl absorption occurred in the autooxidized compound because of formation of the corresponding aldehyde. High resolution MS of the ionic fragments m/e 229, 213 and 212 from (1b) showed that these were due to the loss of $-C_2H_5$, $-3\mathrm{Me}$ and $-\mathrm{Me}\mathrm{-OMe}$, respectively, and therefore were not derived from an aldehyde structure. In the six terpenoid aldehydes that we had previously identified from cotton [9], the C-4 protons occurred at δ 7·39–7·70, and the aldehyde

protons occurred at δ 10·96–11·17. Aldehyde protons generally appear downfield beginning at δ 8·9 [10]. Thus, the proton that others observed at δ 7·3 is the C₄-aromatic proton.

The correct structure of both (1b) and the compound isolated by Zaki et al. [4] appears to be 3-hydroxy-5-isopropyl-4-methoxy-7-methyl-2H-naphtho[1,8-bc] furan. The appropriate trivial name for this compound is desoxy-6-methoxyhemigossypol, because of its relationship to hemigossypol [3], and to desoxygossypol tetramethyl ether that was synthesized by Adams and Dial [11]. The structure of (1b) was confirmed by its photooxidation to 6-methoxyhemigossypol. The structure of (1b) is further supported by its co-occurrence with desoxyhemigossypol (1a) that readily photooxidizes to hemigossypol.

Antimicrobial activity was reported for vergosin [4,5] but the bioassay consisted of direct layering of nutrient agar on TLC plates followed by seeding with fungal spores and incubation for several days. Under these conditions, desoxy-6-methoxyhemigossypol is readily oxidized to 6-methoxyhemigossypol, a known antimicrobial substance [3]. Thus, more careful bioassays are required to evaluate the biological activities of the 2*H*-naphtho[1,8-*bc*]furans.

Compounds (1a) and (1b) accumulate rapidly in cotton tissues 12–96 hr after infection with several fungi (unpublished data). The ready conversion of these compounds to the phytoalexins, hemigossypol and 6-methoxyhemigossypol, by UV-catalyzed oxidation, suggests that they are the natural biological precursors of these phytoalexins. Veech and Bell (unpublished) have recently shown that a peroxidase-type of enzyme converts hemigossypol to gossypol in cotton. Since such conversion normally involves free radical formation, the same or a similar enzyme might convert (1a) and (1b) to their corresponding aldehydes.

EXPERIMENTAL

Crude terpenoid fractions were prepared from stele tissues of Gossypium barbadense L. "Sea Brook Sea Island 12B2" 48 hr after inoculation with Verticillium dahliae (strain 117). Extraction procedures have been described [3,12] and all were conducted in subdued light or in darkness. Chromatography was on polyamide (System 1) and Si gel (Syst. 3–6) as in [3], or Si gel with C_6H_6 . (Syst. 8), and Et_2O —naphtha (1:1), (Syst. 9). TLC plates were irradiated with UV (254 nm) for 1 hr to convert the desoxy terpenoids to their aldehydes which were then detected with 2% phloroglucinol in EtOH—conc HCl (1:1).

Spectra. UV spectra were determined in 95% EtOH or 95% EtOH containing 0·03 M NaOH (EtONa) and are reported as $\lambda_{\rm max}$ (ϵ). IR spectra were determined in KBr. The 100 MHz NMR spectra were determined in CDCl₃ at 23° and are reported in δ units. Hydroxyl protons were detected by exchange with D₂O. MS were obtained by direct probe at 25° with a source temp. of 200°.

Isolation of desoxyhemigossypol (1a) and desoxy-6-methoxyhemigossypol (1b). Compound (1a) was purified from crude terpenoid extracts by TLC with sequential use of systems 4 (R_f 0·22), 3 (R_f 0·39), 1 (R_f 0·33), and 6 (R_f 0·47) given in Ref. [3]. Compound (1b) was purified by use of systems 8 (R_f 0·31), 3 (R_f 0·77), and 4 (R_f 0·41) in sequence as described in Ref. [3]. Compound (1a) crystallized from C_6H_6 -hexane as pale yellow crystals. Mp 134–138°. UV $\lambda_{\rm max}^{\rm FOOH}$ (ε) 224 (31000), 246 (38 800), 251 (sh), 294 (7900), 303 (7800), 331 (sh) nm; $\lambda_{\rm max}^{\rm EEONa}$ (ε) 254 (44700), 260 (sh), 299 (10 800), 310 (10 900), 325 (8800), 338 (8400) nm. IR: $\nu_{\rm max}^{\rm KBF}$ 3400–3300, 1655, 1633, 1505, 1291 cm⁻¹. Compound (1b) also crystallized from C_6H_6 -hexane as pale yellow crystals. Mp 121·5–123°. UV $\lambda_{\rm max}^{\rm EEOH}$ (ε) 224 (39 000), 247 (62 500), 284 (sh), 295 (10000), 306 (10 700), 325 (sh), 339 (5300) nm; $\lambda_{\rm max}^{\rm EEON}$ (ε) 254 (65 700), 290 (sh), 299 (15 000), 311 (16500), 341 (10000) nm. IR: $\nu_{\rm max}^{\rm KBF}$ 3430–3340, 1655, 1642, 1622, 1508 cm⁻¹.

Photooxidation. Pure (1a) and (1b) were developed in TLC system 5 [3], and plates were immediately exposed to UV radiation (both 254 and 366 nm) in air for 1 hr. Oxidation products were eluted from the bands with Et_2O and purified by TLC. The major oxidation product of (1a) was hemigossypol. It was purified by sequential TLC in systems 1 (R_f 0·31), 3 (R_f 0·64), and 4 (R_f 0·41). The major oxidation product of (1b) was 6-methoxygossypol. It was purified in systems 5 (R_f 0·43) and 9 (R_f 0·59). The aldehydes formed by photooxidation were identified by comparing their chromatographic behavior and MS with those of authentic samples.

Acknowledgements—Appreciation is expressed to Ronald Grigsby, Texas A&M University, who performed the high-resolution mass measurements, and to Mildred E. Hummel and Gilbert Tribble for technical assistance.

REFERENCES

- 1. Bell, A. A. (1967) Phytopathology 57, 759.
- 2. Bell, A. A. (1969) Phytopathology 59, 1119.
- 3. Bell, A. A., Stipanovic, R. D., Howell, C. R. and Fryxell, P. A. (1975) *Phytochemistry* 14, 225.
- Zaki, A. I., Keen, N. T., Sims, J. J. and Erwin, D. C. (1972) Phytopathology 62, 1398.
- Zaki, A. I., Keen, N. T. and Erwin, D. C. (1972) Phytopathology 62, 1402.
- Berry, D. and Smith, D. C. C. (1972) J. Chem. Soc. (C) 699
- 7. Stipanovic, R. D., unpublished results.
- 8. Reid, D. H. (1965) Quart. Rev. 19, 274.
- Stipanovic, R. D., Bell, A. A., Mace, M. E. and Howell, C. R. (1974) in *Beltwide Cotton Prod. Res. Conf.*, *Proc.*, Dallas, Texas, pp. 41–45. National Cotton Council, Memphis, Tennessee.
- Jackman, L. M. and Sternhell, S. (1969) in Nuclear Magnetic Resonance in Organic Chemistry, p. 191. Pergamon Press, Oxford.
- Adams, R. and Dial, W. R. (1930) J. Am. Chem. Soc. 61. 2077.
- Bell, A. A., Stipanovic, R. D., Howell, C. R. and Mace, M. E. (1974) in *Beltwide Cotton Prod. Res. Conf., Proc.*, Dallas, Texas, pp. 40-41. National Cotton Council, Memphis, Tennessee.